

Accepted Manuscript

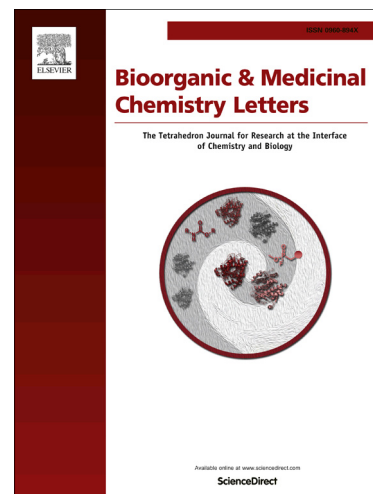
Microwave-Assisted Synthesis, Characterization and Cytotoxic Studies of Novel Estrogen Receptor α Ligands towards Human Breast Cancer Cells

Hanumantharayappa Bharathkumar, Chakrabhavi Dhananjaya Mohan, Hanumappa Ananda, Julian E. Fuchs, Feng Li, Shobith Rangappa, Mohan Surender, Krishna C. Bulusu, Kesturu S. Girish, Gautam Sethi, Andreas Bender, Basappa, Kanchugarakoppal S Rangappa

PII: S0960-894X(15)00042-6
DOI: <http://dx.doi.org/10.1016/j.bmcl.2015.01.030>
Reference: BMCL 22370

To appear in: *Bioorganic & Medicinal Chemistry Letters*

Received Date: 15 October 2014
Revised Date: 31 December 2014
Accepted Date: 17 January 2015



Please cite this article as: Bharathkumar, H., Mohan, C.D., Ananda, H., Fuchs, J.E., Li, F., Rangappa, S., Surender, M., Bulusu, K.C., Girish, K.S., Sethi, G., Bender, A., Basappa, Rangappa, K.S., Microwave-Assisted Synthesis, Characterization and Cytotoxic Studies of Novel Estrogen Receptor α Ligands towards Human Breast Cancer Cells, *Bioorganic & Medicinal Chemistry Letters* (2015), doi: <http://dx.doi.org/10.1016/j.bmcl.2015.01.030>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

**Microwave-Assisted Synthesis, Characterization and Cytotoxic Studies of Novel
Estrogen Receptor α Ligands towards Human Breast Cancer Cells**

Hanumantharayappa Bharathkumar,^a Chakrabhavi Dhananjaya Mohan,^b Hanumappa Ananda,^b Julian E. Fuchs,^c Feng Li,^d Shobith Rangappa,^e Mohan Surender,^f Krishna C. Bulusu,^c Kesturu S. Girish,^g Gautam Sethi,^d Andreas Bender,^c Basappa,^{a,*} Kanchugarakoppal S Rangappa,^{b,*}

^a*Laboratory of Chemical Biology, Department of Chemistry, Bangalore University, Palace Road, Bangalore 560 001, India*

^b*Department of Chemistry, University of Mysore, Mysore 570 006, India*

^c*Centre for Molecular Informatics, Department of Chemistry, University of Cambridge, Lensfield Road, CB2 1EW, Cambridge, United Kingdom*

^d*Department of Pharmacology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore 117 597*

^e*Frontier Research Center for Post-genome Science and Technology Hokkaido University, Sapporo 060 0808 Japan*

^f*CAS in Crystallography and Biophysics, University of Madras, Chennai, Tamil Nadu-600 025, India*

^g*Department of Studies and Research in Biochemistry, Tumkur University, Tumkur 572 103, India*

Abstract: A new, simple, and microwave-assisted, solution-phase T3P[®]-DMSO mediated method for the preparation of a novel class of estrogen receptor alpha (ER α) ligands based on the 2-phenylquinoline scaffold was developed. Furthermore, the novel ER α ligands were tested for their bioactivity against ER α -positive and ER α -negative cell lines. The ligand (entry 4), with amine and nitro group substitution at C4 position, displayed significant cytotoxicity against MCF-7 and HepG2 cells with an IC₅₀ value of 6 and 11 μ M, respectively. On the other hand, ER α -negative cells displayed resistance to quinolines induced cytotoxicity with an IC₅₀ value >100 Mm and they does not induce cytotoxicity in normal breast epithelial cells. Molecular docking analyses suggest a consistent binding mode for these ER α ligands in the ligand binding domain of the human ER α and predict the ligands to occupy the hydrophobic cavity in a similar fashion as estradiol or GW2368.

INTRODUCTION

Breast cancer is a leading cancer in women worldwide and contributing to second cause of lethality after lung cancer.^{1, 2} Most breast cancers are associated with interaction of estrogen receptors (ER) in the breast epithelial cells to estrogen. The physiological action of estrogen is induced via two types of estrogen receptors namely ER α and ER β .³ Research in the previous decade revealed that more than 70% of breast cancers are due to ER α dependent epithelial cell proliferation. The role of ER β is not clear in initiation and progression of breast cancer.⁴ ER α belongs to nuclear receptor superfamily which regulates the transcription of genes involved in proliferation, anti-apoptosis, metastasis and immunosurveillance.⁵⁻⁷ The binding of 17 β -estradiol to ER α induces the receptor dimerization and facilitates binding of the ligand-receptor complex to the promoter of target genes.⁸ Also, ER-dependent pathways regulate the synthesis and distribution of glycosaminoglycans in cancer cells.⁹ Several small molecule ER α antagonists including Tamoxifen, Raloxifene and Fulvestrant have been implemented in the treatment of breast cancer.^{10, 11} Benzisoxazole tethered azoles have known to be the better ER ligands.¹²⁻¹⁴ Therefore, probing small molecules against ER α is considered to be the most attractive therapeutic target to treat breast cancer.¹⁵

Quinoline derivatives are the pharmacologically important heterocycles which have been studied extensively for their anticancer properties. Multiple reports have demonstrated the preparation of quinolines using strong base like tertiary butoxide.^{16, 17} We previously reported the anti-cancer effect of various heterocyclic compounds¹⁸⁻²⁰ and recently reported the solution phase synthesis of 2-amino-chromene-3-carbonitriles from alcohols, malanonitrile and phenols.²¹ Using a similar strategy, herein, we report a simple and efficient method for the preparation of T3P[®]-DMSO mediated 2-phenylquinoline derivatives using amino alcohols and acetophenones under microwave irradiation and evaluated for their cytotoxicity. The newly developed method was generalized using variety of aromatic 2-amino alcohols

and different substituted acetophenones to obtain **1-10** molecules (**Scheme 2**; **Table 1**). Our docking analysis validated the interaction of quinoline derivatives with the estrogen receptor in the similar fashion as estradiol to induce its anticancer effect.

Table 1. The physical characterization of new quinoline-based ER α ligands, whose core scaffold has different substitutions at R¹ to R⁶ positions.

Entry	R ¹	R ²	R ³	R ⁴	R ⁵	R ⁶	Y	mp (°C)
1	H	H	OH	H	H	H	85	52
2	CH ₃	H	Br	H	H	H	89	50
3	H	H	Cl	H	Cl	H	92	70
4	H	Cl	H	H	NH ₂	H	95	60
5	CH ₃	H	Cl	H	Cl	H	90	59
6	H	H	H	H	Br	H	88	63
7	CH ₃	H	H	H	H	H	87	54
8	CH ₃	H	H	H	NH ₂	H	91	67
9	CH ₃	H	OH	H	H	OH	89	54
10	CH ₃	H	H	NO ₂	H	H	93	64

Additionally, all the new molecules were characterized completely using IR, ¹H NMR, ¹³C NMR, and LC-MS spectral analysis (please refer supplementary data).

Further, the library of ER α ligands was tested for its cytotoxicity against ER α -positive, ER α -negative cancer cells and their counterpart non-diseased breast epithelial cells as described previously and detailed methodology is provided in supplementary information.^{22, 23} The results are summarized in Table 2.

Table 2. Cytotoxicity data for the new ER α ligands IC₅₀ against human cancer cells.

Entry	ER α -positive cells		ER α -negative cells		Breast epithelial cells
	MCF-7 (μ M)	HepG2 (μ M)	MDA-MB-231 (μ M)	BT549 (μ M)	MCF-10A (μ M)
1	>50	>50	NT	NT	NT
2	>50	>50	NT	NT	NT
3	25	31.6	> 100	> 100	> 100
4	6	11	> 100	> 100	> 100
5	>50	>50	NT	NT	NT
6	12	9	> 100	> 100	60.3
7	>50	>50	NT	NT	NT
8	34	41	NT	NT	NT
9	28	19	NT	NT	NT
10	23	37	> 100	> 100	> 100

*NT – Not tested

Among the tested compounds **4**, **6**, and **10** significantly inhibited the proliferation of ER α -positive cells. Further, compounds (**4**, **6**, and **10**) with high cytotoxicity were tested against ER α -negative cells. All the ER α -negative cells were resistance to the lead compounds with IC₅₀ values more than 100 μ M. However, the shortlisted three molecules did not induce cytotoxicity on MCF-10A cells up to 72 h at 100 μ M. These results indicate that the newly synthesized ER α ligands are selectively cytotoxic against ER α expressing cancer cells and does not interfere with viability of their counterpart.

Additionally, the structural models for molecular interactions between the newer ER α ligands and the human estrogen receptor were generated using *in silico* docking analysis as described previously and detailed methodology is provided in supplementary information.²⁴⁻²⁶ Docking

was based on the co-crystal structure of the naphthalene derivative GW2368 with the estrogen receptor (PDB: 3DT3).²⁷ Molecular docking suggests a consistent binding mode for the series of quinoline-based ER α ligands in the ligand binding domain of the human estrogen receptor. Thereby, the compounds occupy the hydrophobic cavity in a similar fashion as estradiol or GW2368 (Figure 1). The docking scores of the ER α ligands were comparable with estradiol or GW2368 (Supplementary Table 1). The quinoline scaffold of the ER α ligands occupies the position of rings A and B in the steroid and show major overlap with the naphthalene ring system of GW2368. Furthermore, presence of benzene substituents allows for interactions with His-524 in agreement with other estrogen receptor ligands. Presence of a hydrogen bond donor function is predicted to facilitate a second binding mode, where hydrogen bonds to Arg-394 and Glu-353 are formed. The structure-activity-relationship studies for the compound 4, which bearing chlorine atom at R² renders significant anti-proliferative activity towards ER positive cancer cells, whereas the presence of methyl group at R¹ decreases the activity. This observation was evidenced with strong binding of compound 4 to ER alpha LBD, when compared to compound 8.

In conclusion, we have identified a novel quinoline-based ER α ligands as biologically active compounds against ER α expressing human cancer cells. This study also introduces a novel microwave-assisted synthesis pathway to the compound series. Therefore, method will be useful to develop libraries of quinoline-based ER α ligands to treat breast cancer.

Acknowledgement

This research was supported by University Grants Commission (41-257-2012-SR), Vision Group Science and Technology, Department of Science and Technology (NO.SR/FT/LS-142/2012) to Basappa. This work was supported by NUHS Bench-to- Bedside-To-Product grant to GS. Deanship of Scientific Research, College of Science Research Centre, King Saud

University, Kingdom of Saudi Arabia is also acknowledged. Basappa & CDM thank Karnataka University, India & DST for DC PAVATE & INSPIRE Fellowships.

References and Notes

1. Karadedou, C. T. *Hippokratia* **2006**, *10*, 128.
2. Ma, J.; Jemal, A. In *Breast Cancer Metastasis and Drug Resistance*; Ahmad, A., Ed.; Springer New York, 2013, pp. 1.
3. Ali, S.; Coombes, R. C. *Nat. rev. Cancer* **2002**, *2*, 101.
4. Hall, J. M.; Couse, J. F.; Korach, K. S. *J. Biol. Chem.* **2001**, *276*, 36869.
5. Gradishar, W. J.; Cella, D. *Jama*. **2006**, *295*, 2784.
6. Boonyaratanakornkit, V.; Edwards, D. P. *Essays Biochem.* **2004**, *40*, 105.
7. Smyth, J. F.; Gourley, C.; Walker, G.; MacKean, M. J.; Stevenson, A.; Williams, A. R.; Nafussi, A. A.; Rye, T.; Rye, R.; Stewart, M.; McCurdy, J.; Mano, M.; Reed, N.; McMahon, T.; Vasey, P.; Gabra, H.; Langdon, S. P. *Clin. Cancer Res.* **2007**, *13*, 3617.
8. Mao, C.; Patterson, N. M.; Cherian, M. T.; Aninye, I. O.; Zhang, C.; Montoya, J. B.; Cheng, J.; Putt, K. S.; Hergenrother, P. J.; Wilson, E. M.; Nardulli, A. M.; Nordeen, S. K.; Shapiro, D. J. *J. Biol. Chem.* **2008**, *283*, 12819.
9. Fongmoon, D.; Shetty, A. K.; Basappa; Yamada, S.; Sugiura, M.; Kongtawelert, P.; Sugahara, K. *J. Biol. Chem.* **2007**, *282*, 36895.
10. Fisher, B.; Costantino, J. P.; Wickerham, D. L.; Redmond, C. K.; Kavanah, M.; Cronin, W. M.; Vogel, V.; Robidoux, A.; Dimitrov, N.; Atkins, J.; Daly, M.; Wieand, S.; Tan-Chiu, E.; Ford, L.; Wolmark, N. *J. Natl. Cancer Inst.* **1998**, *90*, 1371.
11. Fabian, C. J.; Kimler, B. F. *J. Clin. Oncol.* **2005**, *23*, 1644.
12. Malamas, M. S.; Manas, E. S.; McDevitt, R. E.; Gunawan, I.; Xu, Z. B.; Collini, M. D.; Miller, C. P.; Dinh, T.; Henderson, R. A.; Keith, J. C., Jr.; Harris, H. A. *J. Med. Chem.* **2004**, *47*, 5021.
13. Rangappa, K. S. *J. Phys. Org. Chem.* **2005**, *18*, 773.
14. Mantelingu, K.; Sadashiva, M.; Rangappa, K. *Indian J. Chem. Sect. B.* **2004**, *43*, 1954.
15. Tiwari, A.; Shivananda, S.; Gopinath, K. S.; Kumar, A. *J. Biol. Chem.* **2014**.
16. V Kouznetsov, V.; A Rojas Ruiz, F.; Y Vargas Mendez, L.; P Gupta, M. *Lett. Drug Des. Discov.* **2012**, *9*, 680.
17. Vu, A. T.; Cohn, S. T.; Manas, E. S.; Harris, H. A.; Mewshaw, R. E. *Bioorg. Med. Chem. Lett.* **2005**, *15*, 4520.
18. Bharathkumar, H.; Paricharak, S.; Dinesh, K.; Siveen, K. S.; Fuchs, J. E.; Rangappa, S.; Mohan, C.; Mohandas, N.; Kumar, A. P.; Sethi, G. *RSC Advances* **2014**, *4*, 45143.
19. Anusha, S.; Anandakumar, B.; Mohan, C. D.; Nagabhushana, G.; Priya, B.; Rangappa, K. S. *RSC Adv.* **2014**, *4*, 52181.
20. Priya, B. S.; Anil Kumar, C.; Nanjunda Swamy, S.; Basappa; Naveen, S.; Shashidhara Prasad, J.; Rangappa, K. S. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 2775.
21. Keerthy, H. K.; Garg, M.; Mohan, C. D.; Madan, V.; Kanojia, D.; Shobith, R.; Nanjundaswamy, S.; Mason, D. J.; Bender, A.; Basappa; Rangappa, K. S.; Koeffler, H. P. *PloS one* **2014**, *9*, e107118.
22. Keerthy, H. K.; Mohan, C. D.; Siveen, K. S.; Fuchs, J. E.; Rangappa, S.; Sundaram, M. S.; Li, F.; Girish, K. S.; Sethi, G.; Basappa, B.; Bender, A.; Rangappa, K. S. *J. Biol. Chem.* **2014**, *289*, 31879.

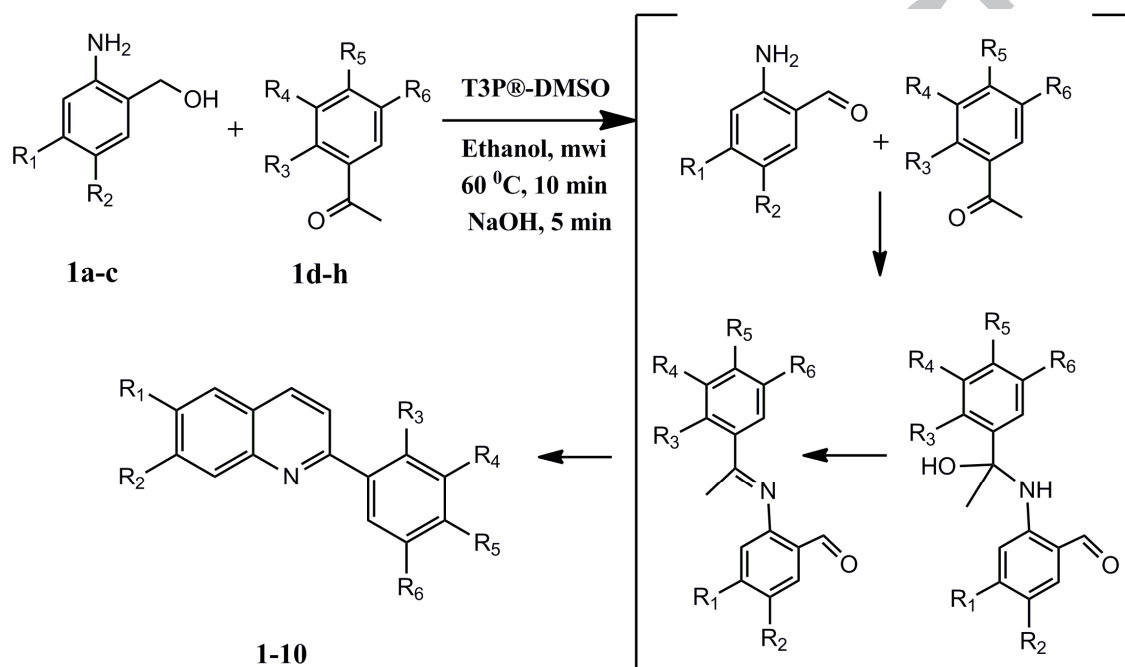
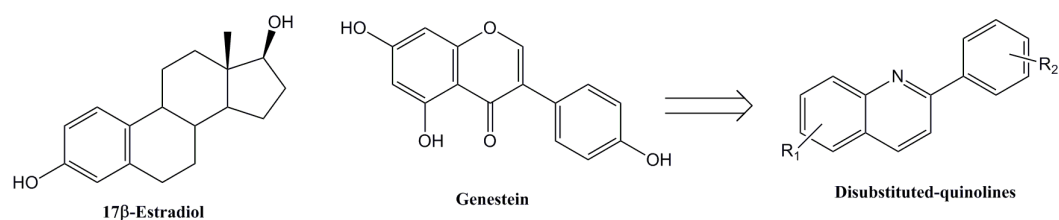
23. Mohan, C. D.; Bharathkumar, H.; Bulusu, K. C.; Pandey, V.; Rangappa, S.; Fuchs, J. E.; Shanmugam, M. K.; Dai, X.; Li, F.; Deivasigamani, A.; Hui, K. M.; Kumar, A. P.; Lobie, P. E.; Bender, A.; Basappa; Sethi, G.; Rangappa, K. S. *J. Biol. Chem.* **2014**, 289, 34296.
24. Labute, P. *Proteins* **2009**, 75, 187.
25. De Lano W: The Pymol Molecular Graphics System, v., De Lano Scientific, San Carlos, CA.
26. Brzozowski, A. M.; Pike, A. C.; Dauter, Z.; Hubbard, R. E.; Bonn, T.; Engstrom, O.; Ohman, L.; Greene, G. L.; Gustafsson, J. A.; Carlquist, M. *Nature* **1997**, 389, 753.
27. Fang, J.; Akwabi-Ameyaw, A.; Britton, J. E.; Katamreddy, S. R.; Navas, F., 3rd; Miller, A. B.; Williams, S. P.; Gray, D. W.; Orband-Miller, L. A.; Shearin, J.; Heyer, D. *Bioorg. Med. Chem. Lett.* **2008**, 18, 5075.

Figure Legends

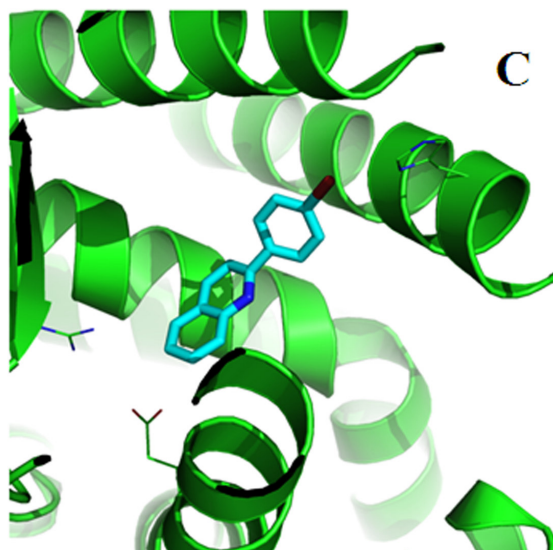
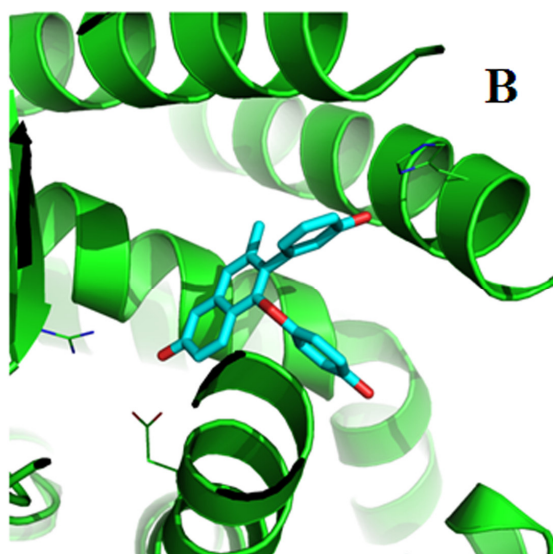
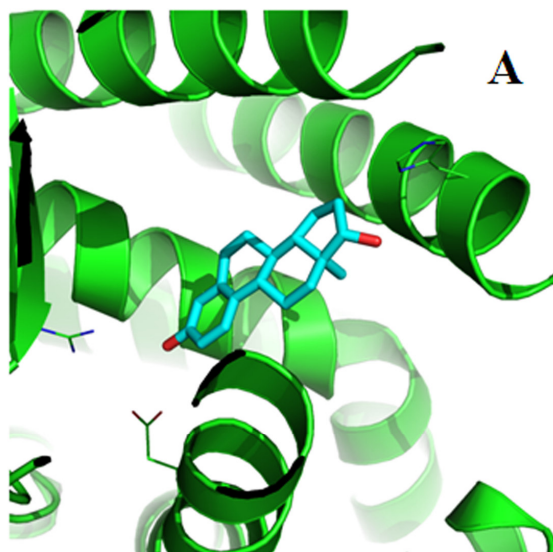
Figure 1: ER α Scaffold Evolution

Figure 2: Schematic representation of the synthesis of quinoline-based ER α ligands.

Figure 3: Predicted interactions of quinoline-based ER α molecules towards the ligand binding domain of the human estrogen receptor. The protein is shown as green cartoon with main polar interaction centers Arg-394, Glu-353, and His-524 highlighted as sticks in atomic coloring. Interactions of estradiol with estrogen receptor (A) are resembled by the naphthalene-derived compound GW2368 (B) and predicted to be similar for the quinoline series (C).



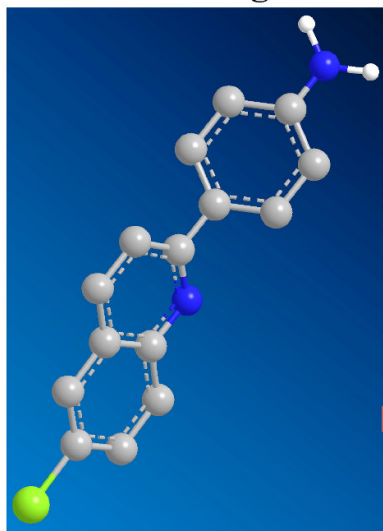
Scheme 2



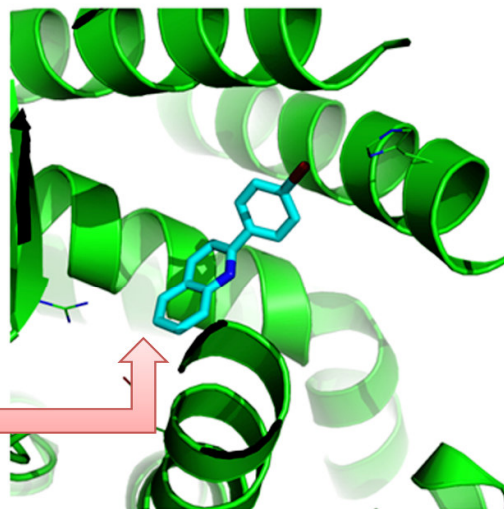
**Microwave-Assisted Synthesis, Characterization and Cytotoxic Studies of Novel
Estrogen Receptor α Ligands towards Human Breast Cancer Cells**

Hanumantharayappa Bharathkumar, Chakrabhavi Dhananjaya Mohan, Hanumappa Ananda,
Julian E. Fuchs, Feng Li, Shobith Rangappa, Mohan Surender, Krishna C. Bulusu, Kesturu S.
Girish, Gautam Sethi, Andreas Bender, Basappa, Kanchugarakoppal S. Rangappa

Lead ER α ligand



MCF-7 cells $IC_{50} = 6 \mu M$



Newer ligands occupy the ligand binding domain of ER α similar to estradiol or GW2368